

# **Diagnostic Exome Sequencing Suggests Digenic Inheritance** in a Cobalamin Metabolism Disorder

#### BACKGROUND

- Diagnostic exome sequencing is being applied clinically resulting in elevated diagnostic yield from genetic testing (~33-50%).
- Exome testing is useful in generating hypotheses for novel disease-associated genes or patterns of inheritance.
- One challenge in DNA-based diagnosis is the availability of an appropriate sample. Genetic disease leading to spontaneous abortion, fetal demise, or early neonatal or childhood death often results in a missed opportunity to collect and store appropriate material for further testing- something that often becomes more valuable to the family as grieving abates and they confront the possibility of recurrence in a subsequent pregnancy.

### **METHODS**

- > Genomic deoxyribonucleic acid (gDNA) was isolated from whole blood from the parents and unaffected sister. Samples were prepared using the SureSelect Target Enrichment System (Agilent Technologies, Santa Clara, CA). The enriched exome libraries were sequenced using paired-end, 100-cycle chemistry on the Illumina HiSeq 2000 (Illumina, San Diego, CA). Several months later, we were notified of the availability of Guthrie blood spot card from the patient and extracted DNA in order to perform single site mutation analysis on positive findings.
- > Exome data undergoes alignment, base calling, and variant calling. Passing base calls have at least 7x coverage and quality scores of Q20 or higher, which translates to a base call error rate probability of 1:100, or a base call read accuracy of 99%. Exons plus at least 2 bases into the 5' and 3' ends of all the introns are analyzed and reported. Variants were filtered further based on family history and possible inheritance models. Data is annotated with the Ambry Variant Analyzer tool (AVA), including nucleotide and amino acid conservation, biochemical nature of amino acid substitutions, population frequency, predicted functional impact, and clinical disease associations (Human Gene Mutation Database (HGMD; Stenson, 2009)), OMIM, and several other databases).
- We sequenced the exome of the parents and unaffected sister of an infant with suspected cobalamin metabolism disorder and for whom no appropriate DNA sample was available for exome sequencing.
- > A molecular geneticist performed interpretive filtering based on the deleterious nature of the candidate alterations literature search and analysis of the relevance of the candidate genes' function in relation to the patient's phenotype.
- > Each candidate variant was analyzed by Sanger sequencing for mutation confirmation and co-segregation analysis.

Figure 1. Family Pedigree			

Stepwise Variant Filtering <sup>1</sup>	Father	ner Mother Sig		
No. of variants in coding regions <sup>2</sup>	110,636 108,825			
No.post-removal of intergenic and 3'/5' UTR variants	82,300 81,626 83,4			
No. post-removal of non-splice-related intronic <sup>3</sup> variants	21,855 21,963 22,31			
No. post-removal of synonymous variants	11,622	11,874		
A) Autosomal recessive (mother and father carrier, daughter negative or carrier)	2 genes (4 alterations)			
No. of genes in A) related to phenotype	0 genes (0 alterations)			
B) X-linked recessive (mother carrier)	1 gene (1 alteration)			
No. of genes in B) related to phenotype	0 genes (0 alterations)			
C) Autosomal recessive carrier variants (mother and father):	58	51	N/A	
No. of genes in C) related to phenotype	2 genes (2 alterations) $^4$			





# **RESULTS/ DISCUSSION**

# Table 2. Coverage of the

#### Table 3. Sanger sequencing confirmation and co-segregation

- Through custom bioinformatics analysis and family history variant filtering, we identified mutations in two genes in the cobalamin metabolism, *LMBRD1* and *MTR* (Figure 1, Table 1).
- Special consideration was given to the known cobalamin metabolism genes. The majority of the exons among these genes were well covered (Table 2). No variations were identified, other than in MTR and LMBRD1.
- LMBRD1 c.1056delG is the most frequently reported alteration, found to be present in 75% (18/24 chromosomes) of one cohort of 12 patients with cblF deficiency (Rutsch, 2011). Likewise, the c.3518C>T (p.P1173L) alteration identified in the *MTR* gene is the most common alteration, observed at a frequency of about 40% (16/38 chromosomes) of patients with cblG deficiency (Watkins, 2002).
- Both alterations are predicted to cause protein damage: The LMBRD1 alteration, c.1056delG (p.L352fsX18), is a truncating mutation translational frameshift with a predicted alternate stop codon; The MTR missense alteration is located at an evolutionarily conserved amino acid (Figure 2).
- The patient's clinical history is consistent with a mixed presentation for cblF and cblG. Biochemical studies also demonstrated overlap between these two subtypes with borderline high levels of methylmalonic acid in blood and urine

Known	Coba	lami	in
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Gene	Gene Coverage (%) <sup>1</sup>				
ABCD4	>99%				
AMN	~55% <sup>2</sup>				
<i>CD320</i>	~80% <sup>3</sup>				
CUBN	>99%				
FTCD	$\sim 77\%^{4}$				
GIF	>99%				
LMBRD1	>99%				
MCEE	>99%				
MMAA	>99%				
MMAB	~91% <sup>5</sup>				
MMACHC	>99%				
MMADHC	>99%				
MTHFR	>99%				
MTR	>99%				
MTRR	>99%				
MUT	>99%				
PCFT	>99%				
TCN1	>99%				
TCN2	>99%				

GENE(S) PROTEIN		<b>REFSEQ ID</b>		ALTERATION	MOTHER	FATHER	DAUGHTER	PATIENT		
	LMBRD1	LMBR1 domain containing 1	NM_018368 NM_000254		c.1056del (p.L352fsX18)	-/-	+/-	+/-	+/-	
	MTR	5-methyltetrahydrofolate- homocysteine methyltransferase			c.3518C>T (p.P1173L)	+/-	-/-	-/-	+/-	
Сс	Figure 3. Cobalamin Metabolic Pathway			➤ H of th	CONCLUSIONS Herein, whole exome sequencing and custom bioinformatics analysis of the parents of a deceased child with cobalamin deficiency revealed that each parent was a carrier of a well-characterized heterozygous					
		Methylmalonic aciduria		m de	nutation within a wo	ell-describe	d gene ass	sociated with	cobalamin	
Mitochondrion Methylmalonyl-CoA mutase-AdoCbl <i>cblA::mut</i> Methylmalonyl-CoA Succinyl-CoA Cytosol AdoCbl				The identified alterations are likely founder mutations among individuals of European ancestry, consistent with the family's repor ancestry herein (Rutsh, 2011, Watkins, 2002).						orte
	Lysosome CD/F TC-CbI-R C	$bI-R$ $\rightarrow$ $Cob(II)$ $cbID$ $cbID-2 (cbIH)$ $bI-R$ $\rightarrow$ $Cob(II)$ $cbID-1$ MeCbI $CH_{3}-THF+$		Ti	he alterations were ne proband, reducin umber variation (Cl owever, the mothe ven coverage within	e present in ng, but not NV) on the e er and fathe n this gene	the comp eliminating correspon r's next-ge compared	ound heteroz g, the likeliho ding allele of en sequencing with other ge	ygous state od for copy each gene. g data show enes. reduc	e in /- /

(mixed), decreased plasma methionine (cblG) and homocysteinuria (cblF) (See Figure 3 for cobalamin metabolic pathway.)

Both parents are carriers of a single mutation. The mother carriers a well described missense mutation in *MTR* (c.3518C>T; p.P1173L), while both the father and unaffected sister carry the LMBRD1 mutation (c.1056delG; p.L352fsX18) (Table 3).

Several months later, we were notified of the availability of Guthrie blood spot card from the patient and confirmed the presence of both mutations in the proband by Sanger sequencing (Table 3).

<sup>1</sup>G ene coverage is based on % of coding base pairs with  $\geq$  10x coverage and quality scores of  $\geq Q30$  $^{2}AMN$  gene low coverage regions include CDS 1, 7, 8, 9, 10, 11  $^{3}CD320$  gene low coverage regions include CDS 1, 2, 3 <sup>4</sup>*FTDC* gene low coverage regions include CDS 3, 4, 9, 10, 12  ${}^{5}MMAB$  gene lowcoverage regions include CDS 6



• Froese DS, et al. (2010) Exp Rev Mol Med **12**(e37):1-20. **REFERENCES:** • Rutsch F, et al. (2011) J Inherit Metab Dis **34**:121-126.

• Watkins D, et al. (2002) Am J Hum Genet **71**:143-153.

the likelihood for the presence of deletion within the gene (~120X vs.mean= 134X+/-44.4). Whole gene deletions of either gene including exon 11 of *LMBRD1* or exon 31 or *MTR* are excluded on the basis of heterozygosity of the mutation call.

Biochemical analysis and *in vitro* studies are ongoing to further define the impact of digenic haploinsufficiency on this metabolic pathway.

This unique case highlights the power of whole exome sequencing as a diagnostic tool even in the absence of a sample from the affected individual, and suggests a novel mechanism and inheritance pattern for disorders of cobalamin metabolism.